the efficacy

of

## Review article

2020 | Volume 6 | Issue 1 | Pages 48-59

#### ARTICLE INFO

## Special Issue: Nanotechnology in Nanomedicine

**Open Access** 

can

immunotherapies

**Received** January 13, 2020 **Revised** March 31, 2020 **Accepted** April 24, 2020

#### \*Corresponding Author

Mohd. Ahmar Rauf

E-mail ahmarrauf2@gmail.com

## Keywords

Glioblastoma (GBM) Neoantigens Immunosurveillance Immunoediting

### How to Cite

Tatiparti K, Rauf MA. Immunoediting and tumor neoantigens that can impact the efficacy of immunotherapies. Biomedical Letters 2020; 6(1):48-59.



Scan QR code to see this publication on your mobile device.



This work is licensed under the Creative Commons Attribution Non-Commercial 4.0 International License.

Katyayani Tatiparti, Mohd. Ahmar Rauf\*

Department of Pharmaceutical sciences, Wayne state University, Detroit, MI, USA

impact

Immunoediting and tumor neoantigens

## Abstract

that

Immunoediting is by far the most serious challenge in the immunotherapies that are on the rise for Glioblastoma Multiforme (GBM). However, it is a double-edged sword because this process can be suppressive of the tumor too. Personalized medicine will succeed only if there are effective methods in place to address this challenge. A thorough understanding of the host immune system and the tumor cells heterogeneity and their mutual interactions is necessary. There is a lot of research being pursued in this direction. So far, the main aspect identified for developing immunotherapy for GBM is finding the balance between the immunoediting and the use of neoantigens for neoantigen-directed T-cell therapies.

## Introduction

## Immunosurveillance

The understanding of the role of immunoediting came into full swing after Brunet and Thomas defined a novel concept called immunosurveillance [1]. They proposed that the host immune cells, especially the T-cells, are responsible for hunting down the tumor cells and destroying them. An overview of this concept is presented in Fig. 1. They also conceived the concept of neoantigens produced by the tumor cells as the driving force for the T-cells for recognition [2]. There are many evidences accumulated in literature over the years to show the presence of immunosurveillance. Tumor infiltrating lymphocytes (TIL) are responsible for the attack and elimination of tumors [3]. An abundance of CD8<sup>+</sup> T cells in tumors is a marker for their prognosis such as in colon cancer [4,5], breast cancer [6], carcinomas [7], melanomas [8], and so on. Further, natural killer (NK) cells inside the tumor cells in high density have also shown that presence of immunosurveillance [9-11]. This can also be proved by the development of tumors in patients that have had renal or cardiac transplants that immunosuppression can promote cancerous cells like the lung cancer, liver cancer, breast cancer, skin cancer and so on [12]. This especially true if the donors have had a history of cancer because their organs might hoard the metastatic tumor cells that have survived the immune system of the host [13].

R.D. Schreiber, et al., have shone light on the theory of immunoediting through their research which showed that (1) immunosuppressed mice developed sarcomas more aggressively than the wild type mice because lack of normal immune responses to IFNy; (2) immunosurveillance was part of the entire immune response to the tumor antigens - they observed that the immunocompetent mice did not develop tumors derived from syngeneic mice were implanted in them; (3) T-cell mediated immune responses, especially CD4 and CD8 T-cells, were in place to protect host from tumors followed by development of immunological memory of the tumor antigens. However, they also found that transfection of major histocompatibility complex (MHC) class I and transporter associated with antigen processing 1 (TAP) genes, which are responsible for antigen presentation proteins in tumor cells, was seen in the tumor helping them escape the immune system of the host. They summarized their research by concluding

that "The immune response thus functions as an effective extrinsic tumor-suppressor system. However, this process also leads to the immunoselection of tumor cells that are more capable of surviving in an immunocompetent host, which explains the apparent paradox of tumor formation in immunologically intact individuals" [14].

# Immunoediting

Over the years of further understanding of this concept. it has been found that the immunosurveillance is a only a part of a bigger umbrella process called immunoediting [15–17]. Immunoediting is a concept where the tumor undergoes modifications in the characteristic immunogenicity in the presence of active immune system of the host that may develop immuneresistant tumor cells. It is dynamic process that has a role in aggressive tumor progression. There are three phases of immunoediting - Elimination, Equilibrium, and Escape. The summary of this concept has been given in Fig. 2 [18].

## Elimination

This phase is sometimes used synonymously with immunosurveillance. Both innate and adaptive immune systems have a major role to play in this phase. There might be a role of the nonimmunogenic factors too in the triggering of this process like the p53 mutations or down-regulation that promotes cancer [3]. Inflammatory cytokines from the tumor cells, macrophages, dendritic cells or the stromal cells activate the effector cells including the NK, NKT,  $\gamma\delta$  T cells and the IL-12 and IFN- $\gamma$ . The tumor cell death induced by the innate immune cells release the tumor antigens (TA) that activate the system [19–21]. adaptive immune Further. maturation of dendritic cells occurs, and they arrive at the tumor draining lymph nodes (TDLNs) presenting the TA to the immune system more that repeats the process described to eliminate the tumor cells in all [22]. There are many immune cells other than these primary cells like the  $T_H 17$  activated by IL-17, IL-17F, IL-21, IL-22 and IL-23 receptor (IL-23R), Regulatory T cells (Treg), CD25<sup>+</sup> (IL-2 receptor alpha chain), T-cell-attracting chemokine CCL5 (also called RANTES), monocyte chemotactic protein 1 (MCP-1), nuclear factor-kB ligand (RANKL), etc. that have a role in the process of elimination [23].

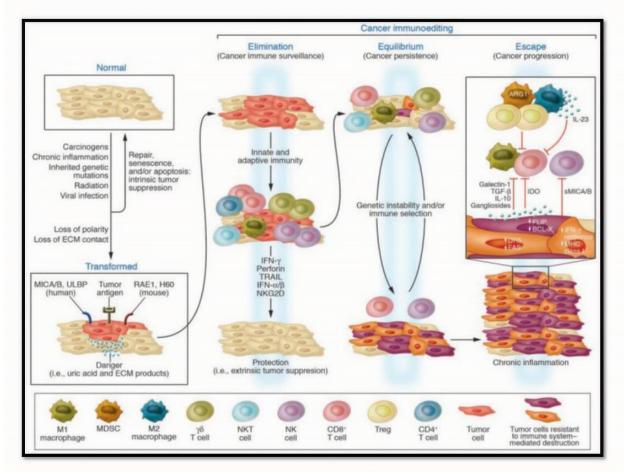


Fig. 1: The overall activity of the immune system in shaping the tumorigenicity

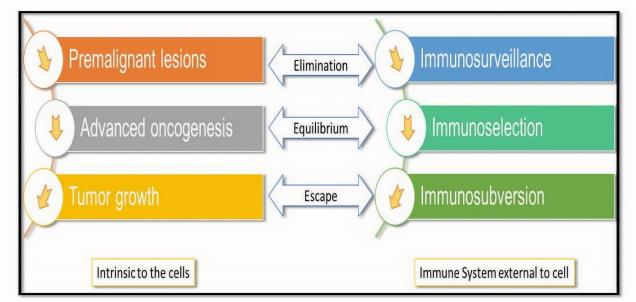


Fig. 2: The three phases of Immunoediting and the key players of the immune system involved

There are four phases to this process:

- **Phase I:** Tumor recognition and cell killing to a small extent occurs when the tumor grows to a discernible size of 2-3 mm that initiates pro-inflammatory responses from the innate immune system [24,25].
- Phase II: Dendritic cells mature and migrate and result in release of IFN-γ that will eventually show the cell killing by antiproliferative [26] and anti-angiogenic effects [27], and induces apoptosis [28]. Further, the necrotic tumor cells are ingested my immature dendritic cells.
- Phase III: IL-12 and IFN- γ produced by NK and macrophages in tumor cells lead to the activation of cytotoxicity mediators like the perforin, TNF-related apoptosis inducing ligand (TRAIL) and reactive oxygen [29,30]. This leads to the presentation of the tumor antigens on their surface to the naïve CD4<sup>+</sup> T cells and the production of the tumor antigen specific CD4<sup>+</sup> and CD8<sup>+</sup> T cells.
- Phase IV: Tumor killing by CD4<sup>+</sup> and CD8<sup>+</sup> T cells is the final step of elimination process by their recruitment at the tumor site producing more IFN- γ. The paradox here is that this is the phase where the tumor cells are educated that reduces their immunogenicity leading to the equilibrium phase of immunoediting [14].

## Equilibrium

This is the next phase of immunoediting process. This phase is responsible for the development of the tumor cells that are resistant to the immune effector cells. There are several hypotheses that explain this phase. One such hypothesis is the absence of inflammatory signals of the immune system to the tumor antigens that activate the dendritic cells and macrophages of the innate immune cascade reactions [14,31–33]. This can lead to tolerance and immune selection of the tumor cells [25]. Another theory explains that the antigens presenting cells sometimes produce anti-inflammatory markers like IL-10 and transforming growth factor- $\beta$  (TGF- $\beta$ ) that fails to elicit immune responses [34]. Yet another theory is the occurrence of random gene mutations promoting tumor [14]. It has also been suggested that the immunogenicity can alter based on the type of cytokines activated mechanism too that can be specific for the type of cancer. Further the immune selection also depends on the immunogenic capabilities of the original mode of tumorigenicity (like the chemical, or viral induction or spontaneous induction) or the original host of the tumor in case of transplantation [35–38] that has a role making the tumor cells more resilient to destruction. This phase is the longest phase of the immunoediting process spanning over years [15].

*Escape:* This phase is where the mutated immune selective cells escape the immune system of the host leading tumor progression. There may be a few ways this happens. The first one may be the alterations developed in the signal transduction molecules of the effector cells. The primary signal transduction cells involved are the T-cell receptors such as TCR-CD3 complex (specifically the CD3- $\zeta$  chain) found on the TILs that are responsible for the expression of TA, Thelper-1 polarization, upregulation of the IL-10 and TGF- $\beta$  and downregulation of IFN- $\gamma$  that elicit the immune responses [39]. A loss of this complex in the tumor cells causes the evasion of the host immune system. This kind of evasion is seen in many cancers like the pancreatic cancer, renal cancer, and several melanomas [40–43]. Further, it has been known that TCR- $\zeta$  is involved in the apoptosis by activating the caspase 3, and downregulation of anti-apoptotic factors like the Bcl-xL and Bcl-2. Thus, the loss of TCR- $\zeta$  can make the T lymphocytes in the tumor vulnerable to apoptosis by increasing the FasL expression on tumor cells [44-47]. This mechanism operates in a vicious cycle. The other way is the emergence of the tumor-derived soluble factors (TDSF) like vascular endothelial growth factor (VEGF), IL-10, TGF-β, prostaglandin E2, soluble phosphatidylserine, soluble Fas, soluble FasL and soluble MHC class I chain-related A (MICA) [48,49,58,50–57] that promote evasion of the immune system of the host by the tumor cells. Moreover, the overall the presentation of the TA may just not be sufficiently enough to activate the immune system efficiently to combat the tumors [59].

# Neoantigens and their impact on the immunotherapies

In the light of these novel understanding of the tumor immunoediting and its interactions with the host immune system, there are a new class of therapeutic agents developed called the neoantigen-directed Tcell based immunotherapy. Neoantigens are the antigens on the tumor cells that are specifically identified by neoantigen-specific T-cell receptors (TCRs). They have known to play an important role in T-cell mediated anti-tumor immunity. There is evidence in the research conducted proving this fact by showing the presence of T-cell recognized antitumor activity against neoantigens, the immune check point blockade being affected by the neoantigens, the rise of the adoptive T-cell based therapies and vaccines developed based on neoantigens [60-63]. While neoantigens are specific tumor cells, there are other tumor antigens called the tumor-associated antigens (TAAs) and cancergermline antigens (CGAs) that are expressed both on healthy as well as well as the tumor cells [64-66]. These two targets of the have shown more tolerance, less affinity (because of their low level of expression), less specific targetability, more chances of autoimmune responses in non-target tissues and thus failure of the therapies based on them in comparison to the ones based on neoantigens [67,68]. Hence, the greatest advantage of neoantigens is that they are identified as true antigens by the immune system that eliminates all the above drawbacks of targeting the TAAs and CGAs [60].

Jiang et al. have very clearly explained the growth of the neoantigens as targets for cancer immunotherapy as shown in **Fig. 3** [69].

There have been several researchers who attempted to show the existence of antitumor immunity after a second challenge by the tumor. All of them have succeeded in proving that the immune system was capable of recognizing and eliminating the tumor cells after the second exposure by malignant cells [70-72]. These findings led to further work in the field to understand that T-cells were involved in the recognition of these tumors and that they were highly specific to the neoantigens [73-75]. They began applying neoantigen reactive T-cells as a part of therapy to achieve maximum regression in melanomas [76,77]. They also found that these therapies were active at high levels in the tumors as well as the blood for a very long time [78]. To maximize the advantages of the neoantigen therapies, several novel techniques are being put into practice to develop personalized medicine/vaccines for cancer treatment like the next-generation sequencing (NGS) technology [79-86]. The most recent developments in this direction are the checkpoint inhibitors like the CTLA-4 and PD-1 inhibitors on the T-cells [87–91] that work by allowing enhanced functioning of the immune responses using these immune checkpoints [92-96]. Like the two sides of a coin, these developments can also be not very effective in the achieving the goals [97] showing that there is still a long road to be explored along [97] and translate them to the clinical settings [98]. The same is true with the developing of vaccines for cancers using the immune checkpoint inhibitors which are on the rise and are actively crossing the pre-clinical barriers in carcinomas melanomas [99–101]. [102,103]. sarcomas [104,105]. Vaccines are also developed for mutation specific antitumor responses. Some of these vaccines have gone to the Phase I clinical trials like the dendritic cell vaccine for melanoma in 2015 [106]; the synthetic long peptide vaccine in conjunction with PD-1 inhibitor for melanomas in 2017 [107]; personalized RNA mutanome vaccine for melanoma combined with PD-1 inhibitor to achieve complete regression of the tumor [108]. Further, glioblastomas are explored now with multiepitope vaccines [109,110] in the Phase I clinical trials. Some of the current clinical trials that are based on neoantigen based vaccines for cancer are listed in Table 1 [69].

Yet another avenue based on neoantigens that is explored now is the adoptive T-cell transfer therapies (ACT) that are tumor neoantigen specific but are manufactured in vitro [62]. These are beneficial in the way that do not have immunosuppressive effects from tumor microenvironment (TME). On the other hand, they also have a disadvantage that it quickly becomes dysfunctional too. They are a very effective in developing personalized therapies for various cancers [111–117]. Needless to say, the ACT also are translated to the clinical trials such as the adoptive transfer ERBB2 interacting protein (ERBB2IP) mutation-reactive  $CD4^+$ tumor infiltrating lymphocytes (TILs) in 2014 for metastatic epithelial cancer, breast cancer [(SLC3A2, KIAA0368, CADPS2, and CTSB)-reactive TILs and colorectal cancer (mutant KRAS G12D reactive CD8<sup>+</sup> TILs) [118–121]. These have shown to achieve reasonable tumor regression. Further, the vaccine and ACT therapies are used in combination with each other and with other conventional therapies or immunotherapies that are already in place for stronger immune responses against tumors [122-131].

# Conclusion

In conclusion, we can say that immunoediting is posing a serious challenge to immune therapies for cancer. However, it is a double-edged sword that not only develops immune-resistant tumor variants but also can be used to our advantage for developing

Data showed that mice could be protected against subsequent re-exposure from tumor cells after surgical removal of the same tumors.		1943 1953	Data suggested that mice were immune against a second challenge with the same carcinogen–induced tumors.
Study reported that antitumor T cells could recognize aberrant peptides derived from tumor– specific mutations.		1988	Neoantigens derived from somatic mutations were identified in various human tumors including melanoma and renal cell carcinoma.
Complete regression in a melanoma patient after infusion of a cell product with a high proportion of neoantigen– reactive T cells.		1996 2004	T cells of the patient with melanoma were reactive against five mutated peptides resulting from somatic point mutations.
Neoantigen-specific T cells	1 -	2005	
could persist at high levels in both the tumor and peripheral blood one month after adoptive transfer.		2012	Firstly applied NGS technology to identify immunogenic neoantigens in mouse tumor models and the protective effects of neoantigen vaccines in B16 tumor model.
found to correlate with		2014	
response to anti-CTLA-4 therapy in melanoma patients.		2014	Neoantigen burden is correlated with response to
The antitumor effect of immune checkpoint blockade was associated with the neoantigen clonality and first mass		2016	anti-PD1 in lung cancer and neoantigen vaccines induce and increase neoantigen specific immunity in melanoma.
spectroscopy identification of T		2017	
cell–recognized neoantigens on human tumor tissue. Neoantigen-directed immune escape in lung cancer evolution.		2019	Personalized neoantigen peptide and RNA vaccines could induce specific T cell populations that recognize autologous tumor.

Fig. 3: History of the tumor neoantigens as immunotherapy targets

#### Biomedical Letters 2020; 6(1):48-59

**Table 1:** List of current clinical trials for neoantigen based vaccines

Interventions	NCT number	Phase	Enrollment status	Cancer types	Combinations
Neoantigen vaccine	NCT03558945	Ι	Recruiting	Pancreatic tumor	None
Neoantigen vaccine	NCT03359239	Ι	Recruiting	Urothelial/bladder cancer	Atezolizumab
Neoantigen vaccine	NCT03645148	Ι	Recruiting	Pancreatic cancer	GM-CSF
Peptide vaccine	NCT03558945	II	Not yet recruiting	TNBC	Nab-paclitaxel, Durvalumab
Peptide vaccine	NCT03929029	Ι	Recruiting	Melanoma	Nivolumab, ipilimumab
Peptide vaccine	NCT03715985	Ι	Recruiting	Solid tumors	None
Peptide vaccine	NCT01970358	Ι	Active, not recruiting	Melanoma	None
Peptide vaccine	NCT03639714	I/II	Recruiting	Solid tumors	Nivolumab, ipilimumab
Peptide vaccine	NCT03956056	Ι	Not yet recruiting	Pancreatic cancer	Adjuvant chemotherapy
Peptide vaccine	NCT02287428	Ι	Active, not recruiting	Glioblastoma	Radiation therapy
Peptide vaccine	NCT02950766	Ι	Recruiting	Kidney cancer	Ipilimumab
Peptide vaccine	NCT03219450	Ι	Not yet recruiting	Lymphocytic leukemia	Cyclophosphamide
Peptide vaccine	NCT03422094	Ι	Recruiting	Glioblastoma	Nivolumab, ipilimumab
DC vaccine	NCT03871205	Ι	Not yet recruiting	Lung cancer	None
DC vaccine	NCT02956551	Ι	Recruiting	NSCLC	None
DC vaccine	NCT03674073	Ι	Recruiting	Hepatocellular carcinoma	Microwave ablation
DC vaccine	NCT03300843	II	Recruiting	Solid tumors	None
RNA vaccine	NCT03908671	Not Applicable	Not yet recruiting	Esophageal cancer, NSCLC	None
RNA vaccine	NCT03480152	I/II	Recruiting	Solid tumors	None
RNA vaccine	NCT03468244	Not Applicable	Recruiting	Solid tumors	None
DNA vaccine	NCT03532217	Ι	Recruiting	Prostate cancer	Nivolumab, Ipilimumab
DNA vaccine	NCT03122106	Ι	Recruiting	Pancreatic cancer	Adjuvant chemotherapy
DNA vaccine	NCT03199040	Ι	Recruiting	TNBC	Durvalumab

novel therapies based on the neoantigens produced during the immunoediting. This is the age of rapidly developing neoantigen based therapies and vaccines and their combinations thereof. These novel strategies of immune therapy that make use of the checkpoint inhibitors must be explored more aggressively while considering the possibility of developing resistance. A thorough understanding and organized extensive research in this area will light up a bright future in the field of personalized immunotherapy.

# **Conflict of interest**

The authors have no affiliations with or involvement in any organization or entity with any financial interest or non-financial interest in the subject matter or materials discussed in this manuscript.

## References

- [1] Burnet M. Cancer; a biological approach. I. The processes of control. Br Med J 1957;1:779–786.
- [2] Thomas L. Cellular and Humoral Aspects of the Hypersensitive States. A Symp New York Acad Med JAMA 1959;170:883.
- [3] Kim R, Emi M, Tanabe K. Cancer immunoediting from immune surveillance to immune escape. Immunology 2007;121:1–14. https://doi.org/10.1111/j.1365-2567.2007.02587.x.
- [4] Stra J, Hinz U, Hasel C, Bhanot U, Mechtersheimer G, Lehnert T, et al. Impaired CD95 expression predisposes for recurrence in curatively resected colon carcinoma: clinical evidence for immunoselection and

CD95L mediated control of minimal residual disease. Gut 2005;54:661–5. https://doi.org/10.1136/gut.2004.052696.

- [5] Naito Y, Saito K, Shiiba K, Ohuchi A, Saigenji K, Nagura H, et al. Advances in Brief CD8 + T Cells Infiltrated within Cancer Cell Nests as a Prognostic Factor in Human Colorectal Cancer o. Cancer Res 1998;58:3491–4.
- [6] Yoshimoto M, Sakamoto G, Ohashi Y, Ph D. Time Dependency of the Influence of Prognostic Factors on Relapse in Breast Cancer. Cancer 1993;72:2993–3001.
- [7] Reichert TE, Day R, Wagner EM, Whiteside TL. Absent or Low Expression of the £ Chain in T Cells at the Tumor Site Correlates with Poor Survival in Patients with Oral Carcinomal. Cancer Res 1998;58:5344-7.
- [8] Haanen JBAG, Baars A, Gomez R, Weder P, Smits M, de Gruijl TD, et al. Melanoma-specific tumorinfiltrating lymphocytes but not circulating melanomaspecific T cells may predict survival in resected advanced-stage melanoma patients. Cancer Immunol Immunother 2006;55:451–8.
- [9] Ishigami S, Natsugoe S, Tokuda K, Nakajo A, Che X, Iwashige H, et al. Prognostic value of intratumoral natural killer cells in gastric carcinoma. Cancer 2000;88:577–83.
- [10] Kondo E, Koda K, Takiguchi N, Oda K, Seike K, Ishizuka M. Preoperative Natural Killer Cell Activity as a Prognostic Factor for Distant Metastasis following Surgery for Colon Cancer. Dig Surg 2003;20:445–451.
- [11] Villegas FR, Cocab S, Villarrubiac VG, Jiménezd R, Chillóna MJ, Jareñoa J, et al. Prognostic significance of tumor infiltrating natural killer cells subset CD57 in patients with squamous cell lung cancer. Lung Cancer 2002;35:23–8.
- [12] Pham SM, Kormos RL, Landreneau RJ, Kawai A, Gonzalez-cancel I, Hardesty RL, et al. Solid Tumors

After Heart Transplantation: Lethality of Lung Cancer. Ann Thorac Surg 1995;60:1623–6.

- [13] Rama I, Grinyó J. Malignancy after renal transplantation: the role of immunosuppression. Nat Rev Nephrol 2010;6:511–9.
- [14] Shankaran V, Ikeda H, Bruce AT, White JM, Swanson PE, Old LJ, et al. IFN $\gamma$  and lymphocytes prevent primary tumour development and shape tumour immunogenicity. Nature 2001;410:1107–1111.
- [15] Dunn GP, Old LJ, Schreiber RD. The Three Es of Cancer Immunoediting. Annu Rev Immunol 2004;22:329–60.
- [16] Dunn G, Koebel C, Schreiber R. Interferons, immunity and cancer immunoediting. Nat Rev Immunol 2006;6:836–48.
- [17] Dunn G, Bruce A, Ikeda H, et al. Cancer immunoediting: from immunosurveillance to tumor escape. Nat Immunol 2002;3:991–8.
- [18] Swann JB, Smyth MJ. Immune surveillance of tumors. J Clin Invest 2007;117:1137–46. https://doi.org/10.1172/JCI31405.antigens.
- [19] Mori S, Jewett A, Murakami-Mori K, Cavalcanti M, Bonavida B. The participation of the Fas-mediated cytotoxic pathway by natural killer cells is tumor-celldependent. Cancer Immunol Immunother 1997;44:282–90.
- [20] Smyth BMJ, Thia KYT, Street SEA, Macgregor D, Godfrey DI, Trapani JA. Perforin-mediated Cytotoxicity Is Critical for Surveillance of Spontaneous Lymphoma. J Exp Med 2000;192:755– 60.
- [21] Takeda K, Hayakawa Y, Smyth MJ, Kayagaki N, Yamaguchi N, Kakuta S, et al. Involvement of tumor necrosis factor-related apoptosis-inducing ligand in surveillance of tumor metastasis by liver natural killer cells. Nat Med 2001;7:94–100.
- [22] Zitvogel L, Terme M, Borg C, Trinchieri G. Dendritic Cell-NK Cell Cross-Talk: Regulation and Physiopathology. Curr Top Microbiol Immunol 2006;298:157–74.
- [23] Bodduluru NL, Kasala ER, Saladi S, Sistla R. Immune system: A double-edged sword in cancer. Inflamm Res 2013;62:823–34. https://doi.org/10.1007/s00011-013-0645-9.
- [24] Matzinger P. Tolerance, Danger, and the Extended Family. Annu Rev Immunol 1994;12:991–1045.
- [25] Smyth M, Godfrey D, Trapani J. A fresh look at tumor immunosurveillance and immunotherapy. Nat Immunol 2001;2:293–299.
- [26] Gollob JA, Sciambi CJ, Huang Z, Dressman HK. Gene Expression Changes and Signaling Events Associated with the Direct Antimelanoma Effect of IFN-; Cancer Res 2005;65:8869–78. https://doi.org/10.1158/0008-5472.CAN-05-1387.
- [27] Cells CDT, Qin Z, Schwartzkopff J, Pradera F, Kammertœns T, Seliger B, et al. A Critical Requirement of Interferon □ -mediated Angiostasis for Tumor Rejection. CANCER Res 2003;63:4095– 100.
- [28] Wall L, Burke F, Barton C, Smyth J, Balkwill F. IFN-□ Induces Apoptosis in Ovarian Cancer Cells in Vivo and. Clin Cancer Res 2003;9:2487–96.

- [29] Ikeda H, Old LJ, Schreiber RD. The roles of IFNγ in protection against tumor development and cancer immunoediting. Cytokine Growth Factor Rev 2002;13:95–109.
- [30] Sinha P, Clements VK, Miller S, Ostrand-Rosenberg S. Tumor immunity: a balancing act between T cell activation, macrophage activation and tumor-induced immune suppression. Cancer Immunol Immunother 2005;54:1137–1142.
- [31] Svane IM, Engel A, Nielsen M, Ljunggren H, Rygaard J, Werdelin O. Chemically induced sarcomas from nude mice are more immunogenic than similar sarcomas from congenic normal mice. Eur J Immunol 1996;26:1844–50.
- [32] Engel A, Svane I, Rygaard J, Werdelin O. MCA sarcomas induced in scid mice are more immunogenic than MCA sarcomas induced in congenic, immunocompetent mice. Scand J Immunol 1997;45:463–70.
- [33] Smyth BMJ, Thia KYT, Street SEA, Cretney E, Trapani JA, Taniguchi M, et al. Differential Tumor Surveillance by Natural Killer (NK) and NKT Cells. J Exp Med 2000;191:661–8.
- [34] Kim R, Emi M, Tanabe K. Cancer cell immune escape and tumor progression by exploitation of antiinflammatory and pro- inflammatory responses. Cancer Biol Ther 2005;4:924–33. https://doi.org/10.4161/cbt.4.9.2101.
- [35] Cankovic M, Linden MD, Zarbo RJ. Use of Microsatellite Analysis in Detection of Tumor Lineage as a Cause of Death in a Liver Transplant Patient. Arch Pathol Lab Med 2006;130:529–32.
- [36] Detry O, Roover A De, Leval L De, Herens C, Delwaide J, Honore P. Transmission of an Undiagnosed Sarcoma to Recipients of Kidney and Liver Grafts Procured in a Non-Heart Beating Donor. Liver Transplant 2005;11:696–9. https://doi.org/10.1002/lt.20457.
- [37] Morath C, Schwenger V, Rohmeiss P, Waldherr R, Ritz E, Zeier M, et al. Transmission of Donor-Derived Small-Cell Carcinoma Cells by a Nontumor-Bearing Allograft. Transplantation 2005;80:540–2.
- [38] MacKie R, Reid R, Junor B. Fatal Melanoma Transferred in a Donated Kidney 16 Years after Melanoma Surgery. N Engl J Med 2003;348:567–8.
- [39] Jhnk C, Henne-bruns D. Systemic and Local Immunosuppression in Pancreatic Cancer Patients 1. Clin Cancer Res 2001;7:925s-933s.
- [40] Schmielau J, Nalesnik M, Finn O. Suppressed T-cell receptor zeta chain expression and cytokine production in pancreatic cancer patients. Clin Cancer Res 2001;7:933s-939s.
- [41] Lockhart DC, Chan AK, Mak S, Joo H-G, Daust HA, Carritte A, et al. Loss of T-cell receptor-CD3ζ and Tcell function in tumor-infiltrating lymphocytes but not in tumor-associated lymphocytes in ovarian carcinoma. Surgery 2001;129:749–56.
- [42] Riccobon A, Gunelli R, Ridolfi R, Paola F De, Flamini E, Fiori M, et al. Immunosuppression in Renal Cancer: Differential Expression of Signal Transduction Molecules in Tumor-Infiltrating, Near-

Tumor Tissue, and Peripheral Blood Lymphocytes. Cancer Invest 2004;22:871–7.

- [43] Staibano S, Mascolo M, Tranfa F, Salvatore G, Mignogna C, Bufo P, et al. Tumor infiltrating lymphocytes in uveal melanoma: a link with clinical behavior? Int J Immunopathol Pharmacol 2006;19:171–9.
- [44] Agrawal S, Marquet J, Copie-bergman C, Jouault H, Reyes F, Bensussan A, et al. CD3 Hyporesponsiveness and In Vitro Apoptosis Are Features of T Cells from Both Malignant and Nonmalignant Secondary Lymphoid Organs. J Clin Invest 1998;102:1715–23.
- [45] Gastman BR, Johnson DE, Whiteside TL, Rabinowich H. Advances in Brief Caspase-mediated Degradation of T-Cell Receptor □ -Chain 1. Cancer Res 1999:59:1422–7.
- [46] Kim JW, Wieckowski E, Taylor DD, Reichert TE, Watkins S, Whiteside TL. Fas Ligand – Positive Membranous Vesicles Isolated from Sera of Patients with Oral Cancer Induce Apoptosis of Activated T Lymphocytes. Clin Cancer Res 2005;11:1010–20.
- [47] Gastman B, Johnson D, Whiteside T, Rabinowich H. Tumor-induced apoptosis of T lymphocytes: elucidation of intracellular apoptotic events. Blood 2000;95:2015–23.
- [48] Urosevic M, Dummer R. HLA-G and IL-10 expression in human cancer-different stories with the same message. Semin Cancer Biol 2003;13:337–42.
- [49] Erdoğan B, Uzaslan E, Budak F, Karadağ M, Ediger D, Oral B, et al. The evaluation of soluble Fas and soluble Fas ligand levels of bronchoalveolar lavage fluid in lung cancer patients. Tuberk Toraks 2005;53:127–31.
- [50] Holdenrieder S, Stieber P, Peterfi A, Nagel D, Steinle A, Salih HR. Soluble MICB in malignant diseases: analysis of diagnostic significance and correlation with soluble MICA. Cancer Immunol Immunother 2006;55:1584–1589.
- [51] Bellamy WT, Richter L, Sirjani D, Roxas C, Glinsmann-Gibson B, Frutiger Y, et al. Vascular endothelial cell growth factor is an autocrine promoter of abnormal localized immature myeloid precursors and leukemia progenitor formation in myelodysplastic syndromes. Blood 2001;97:1427–34.
- [52] Munn DH, Sharma MD, Lee JR, Jhaver KG, Johnson TS, Keskin DB, et al. Potential Regulatory Function of Human Dendritic Cells Expressing Indoleamine 2,3-Dioxygenase. Science (80-) 2002;297:1867–70.
- [53] Gabrilovich D, Ishida T, Oyama T, Ran S, Kravtsov V, Nadaf S, et al. Vascular endothelial growth factor inhibits the development of dendritic cells and dramatically affects the differentiation of multiple hematopoietic lineages in vivo. Blood 1998;92:4150– 66.
- [54] Beck C, Schreiber H, Rowley DA. Role of TGF- □ in Immune-Evasion of Cancer. Microsc Res Tech 2001;52:387–95.
- [55] He X, Stuart JM. Prostaglandin E 2 Selectively Inhibits Human CD4 + T Cells Secreting Low Amounts of Both IL-2 and IL-4. J Immunol 1999; 1999;163:6173–9.

- [56] Sinha P, Clements VK, Ostrand-rosenberg S. Interleukin-13 – regulated M2 Macrophages in Combination with Myeloid Suppressor Cells Block Immune Surveillance against Metastasis. Cancer Res 2005;65:11743–52. https://doi.org/10.1158/0008-5472.CAN-05-0045.
- [57] Oyama T, Ran S, Ishida T, Nadaf S, Kerr L, Carbone DP, et al. Vascular Endothelial Growth Factor Affects Dendritic Cell Maturation Through the Inhibition of Nuclear Factor- κ B Activation in Hemopoietic Progenitor Cells. J Immunol 1998; 1998;160:1224–32.
- [58] Emi M, Tanabe K, Ph D, Uchida Y, Toge T, Ph D. The Role of Fas Ligand and Transforming Growth Factor □ in Tumor Progression Molecular Mechanisms of Immune Privilege via Fas-Mediated Apoptosis and Potential Targets for Cancer Therapy. Cancer 2004;100:2281–91. https://doi.org/10.1002/cncr.20270.
- [59] OCHSENBEIN AF, KLENERMAN P, KARRER U, LUDEWIG B, PERICIN M, HENGARTNER H, et al. Immune surveillance against a solid tumor fails because of immunological ignorance. Proc Natl Acad Sci USA 1999;96:2233–8.
- [60] Schumacher T, Scheper W, Kvistborg P. Cancer Neoantigens. Annu Rev Immunol 2019;37:173–200.
- [61] Chu Y, Liu Q, Wei J, Liu B. Personalized cancer neoantigen vaccines come of age. Theranostics 2018;8:4238–46. https://doi.org/10.7150/thno.24387.
- [62] Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. Science (80-) 2015;348:69–74.
- [63] Yarchoan M, Lutz ER, Laheru DA, Elizabeth M. Targeting neoantigens to augment antitumour immunity. Nat Rev Cancer 2017;17:209–22. https://doi.org/10.1038/nrc.2016.154.Targeting.
- [64] Ward JP, Gubin MM, Schreiber RD, States U. The Role of Neoantigens in Naturally Occurring and Therapeutically Induced Immune Responses to Cancer. Adv Immunol 2016;130:25–74. https://doi.org/10.1016/bs.ai.2016.01.001.The.
- [65] Simpson AJG, Caballero OL, Jungbluth A, Chen Y-T, Old LJ. Cancer/testis antigens, gametogenesis and cancer. Nat Rev Cancer 2005;5:615–625.
- [66] Kalejs M, Erenpreisa J. Cancer/testis antigens and gametogenesis: a review and "brain-storming" session. Cancer Cell Int 2005;5:1–11. https://doi.org/10.1186/1475-2867-5-4.
- [67] Coulie PG, Eynde BJ Van den, Bruggen P van der, Boon T. Tumour antigens recognized by T lymphocytes: at the core of cancer immunotherapy. Nat Rev Cancer 2014;14:135–146.
- [68] Pan R, Chung W, Chu M, Chen S, Chen H, Zheng L, et al. Review Article Recent Development and Clinical Application of Cancer Vaccine: Targeting Neoantigens. J Immunol Res 2018;2018:4325874. https://doi.org/10.1155/2018/4325874.
- [69] Jiang T, Shi T, Zhang H, Hu J, Song Y, Wei J, et al. Tumor neoantigens : from basic research to clinical applications. J Hematol Oncol 2019;12:1–13.
- [70] Waldmann TA. Immunotherapy: past, present and future. Nat Med 2003;9:269–277.

- [71] Foley EJ. Antigenic Properties of Methylcholanthreneinduced Tumors in Mice of the Strain of Origin. Cancer Res 1953;13:835–837.
- [72] Gross L. Intradermal Immunization of C3H Mice against a Sarcoma That Originated in an Animal of the Same Line. Cancer Res 1943;3:326–333.
- [73] Daniel Braindle, Brasseur F, Weynants P, Boon T, Eynde BJ Van den. A Mutated HLA-A2 Molecule Recognized by Autologous Cytotoxic T Lymphocytes on a Human Renal Cell Carcinoma. J Exp Med 9 1996;183:2501–8.
- [74] Lobbins PF, El-Gamil M, Li YF, Kawakami Y, Loftus D, Appella E, et al. A Mutated f3-Catenin Gene Encodes a Melanoma-specific Antigen Recognized by Tumor Infiltrating Lymphocytes. J Exp Med 1996;183:1185–92.
- [75] Plaen EDE, Lurquin C, Pel AVAN, Mariam B, Szikora J, Wolfel T, et al. Immunogenic (tum-) variants of mouse tumor P815: Cloning of the gene of tum- antigen P91A and identification of the. Proc Nati Acad Sci USA 1988;85:2274–8.
- [76] Lennerz V, Fatho M, Gentilini C, Frye RA, Lifke A, Ferel D, et al. The response of autologous T cells to a human melanoma is dominated by mutated neoantigens. PNAS 2005;102:16013–8.
- [77] Huang J, El-gamil M, Dudley ME, Yong F, Rosenberg SA, Robbins PF. T Cells Associated with Tumor Regression Recognize Frameshifted Products of the CDKN2A Tumor Suppressor Gene Locus and a Mutated HLA Class I Gene Product. J Immunol 2004; 2004;172:6057–64.

https://doi.org/10.4049/jimmunol.172.10.6057.

- [78] Zhou J, Dudley M, Rosenberg S, Robbins P. Persistence of Multiple Tumor-Specific T-Cell Clones Is Associated with Complete Tumor Regression in a Melanoma Patient Receiving Adoptive Cell Transfer Therapy. J Immunother 2005;28:53–62.
- [79] Castle JC, Kreiter S, Diekmann J, L€ower M, Roemer N van de, Graaf J de, et al. Exploiting the Mutanome for Tumor Vaccination. Cancer Res 2012;72:1081–92. https://doi.org/10.1158/0008-5472.CAN-11-3722.
- [80] Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science (80-) 2015;348:124–8.
- [81] Stevanović S, Pasetto A, Helman SR, Gartner JJ, Prickett TD, Howie B, et al. Landscape of immunogenic tumor antigens in successful immunotherapy of virally induced epithelial cancer. Science (80-) 2017;356:200–5.
- [82] Verdegaal EME, Miranda NFCC de, Visser M, Harryvan T, Buuren MM van, Andersen RS, et al. Neoantigen landscape dynamics during human melanoma-T cell interactions. Nature 2016;536:91-5.
- [83] Tran E, Robbins PF, Lu Y-C, Prickett TD, Gartner JJ, Jia L, et al. T-Cell Transfer Therapy Targeting Mutant KRAS in Cancer. N Engl J Med 2016;375:2255–62.
- [84] Linnemann C, Buuren MM van, Bies L, Verdegaal EME, Schotte R, Calis JJA, et al. High-throughput epitope discovery reveals frequent recognition of neoantigens by CD4+ T cells in human melanoma. Nat Med 2015;21:81–85.

- [85] Robbins PF, Lu Y-C, El-Gamil M, Li YF, Gross C, Gartner J, et al. Mining exomic sequencing data to identify mutated antigens recognized by adoptively transferred tumor-reactive T cells. Nat Med 2013;19:747–752.
- [86] Matsushita H, Vesely MD, Koboldt DC, Rickert CG, Uppaluri R, Magrini VJ, et al. Cancer exome analysis reveals a T-cell-dependent mechanism of cancer immunoediting. Nature 2012;482:400–4.
- [87] Yarchoan M, Hopkins A, Jaffee EM. Tumor Mutational Burden and Response Rate to PD-1 Inhibition. N Engl J Med2 2017;377:2500–1.
- [88] Allen EM Van, Miao D, Schilling B, Shukla SA, Blank C, Zimmer L, et al. Genomic correlates of response to CTLA-4 blockade in metastatic melanoma. Science (80-) 2015;350:207–12.
- [89] Snyder A, Makarov V, Merghoub T, Yuan J, Zaretsky JM, Desrichard A, et al. Genetic Basis for Clinical Response to CTLA-4 Blockade in Melanoma. N Engl J Med2 2014;371:2189–99.
- [90] Chen DS, Mellman I. Elements of cancer immunity and the cancer-immune set point. Nature 2017;541:321–330.
- [91] Chen DS, Mellman I. Oncology Meets Immunology : The Cancer-Immunity Cycle. Immunity 2013;39:1–10. https://doi.org/10.1016/j.immuni.2013.07.012.
- [92] Łuksza M, Riaz N, Makarov V, Balachandran VP, Hellmann MD, Solovyov A, et al. A neoantigen fitness model predicts tumour response to checkpoint blockade immunotherapy. Nature 2017;551:517–520.
- [93] Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. Science (80-) 2017;357:409–13.
- [94] Chalmers ZR, Connelly CF, Fabrizio D, Gay L, Ali SM, Ennis R, et al. Analysis of 100, 000 human cancer genomes reveals the landscape of tumor mutational burden. Genome Med 2017;9:1–14. https://doi.org/10.1186/s13073-017-0424-2.
- [95] Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SAJR, Behjati S, Biankin A V., et al. Signatures of mutational processes in human cancer. Nature 2013;500:415–21. https://doi.org/10.1038/nature12477

https://doi.org/10.1038/nature12477.

- [96] Anagnostou V, Smith KN, Forde PM, Niknafs N, Bhattacharya R, White J, et al. Evolution of Neoantigen Landscape during Immune Checkpoint Blockade in Non – Small Cell Lung Cancer. CANCER Discov 2017;7:264–77. https://doi.org/10.1158/2159-8290.CD-16-0828.
- [97] McGranahan N, Furness AJS, Rosenthal R, Ramskov S, Lyngaa R, Saini SK, et al. Clonal neoantigens elicit T cell immunoreactivity and sensitivity to immune checkpoint blockade. Science (80-) 2016;351:1463– 70.
- [98] Popovic A, Jaffee EM, Zaidi N, Popovic A, Jaffee EM, Zaidi N. Emerging strategies for combination checkpoint modulators in cancer immunotherapy Find the latest version: Emerging strategies for combination checkpoint modulators in cancer immunotherapy. J Clin Invest 2018;128:3209–18.

- [99] Kreiter S, Vormehr M, Roemer N van de, Diken M, Löwer M, Diekmann J, et al. Mutant MHC class II epitopes drive therapeutic immune responses to cancer. Nature 2015;520:692–6.
- [100] Aurisicchio L, Salvatori E, Lione L, Bandini S, Pallocca M, Maggio R, et al. Poly-specific neoantigentargeted cancer vaccines delay patient derived tumor growth. J Exp Clin Cancer Res 2019;38:1–13.
- [101] Duperret EK, Perales-Puchalt A, Stoltz R, G.H. H, Mandloi N, Barlow J, et al. A Synthetic DNA, Multi-Neoantigen Vaccine Drives Predominately MHC Class I CD8+ T-cell Responses, Impacting Tumor Challenge. Cancer Immunol Res 2019;7:174– 182.
- [102] Forghanifard MM, Gholamin M, Moaven O, Farshchian M, Ghahraman M, Aledavood A, et al. Neoantigen in esophageal squamous cell carcinoma for dendritic cell-based cancer vaccine development. Med Oncol 2014;31:191.
- [103] Kuai R, Ochyl LJ, Bahjat KS, Schwendeman A, Moon JJ. Designer vaccine nanodiscs for personalized cancer immunotherapy. Nat Mater 2017;16:489–96.
- [104] Schumacher T, Bunse L, Pusch S, Sahm F, Wiestler B, Quandt J, et al. A vaccine targeting mutant IDH1 induces antitumour immunity. Nature 2014;512:324–7.
- [105] Gubin MM, Zhang X, Schuster H, Caron E, Ward JP, Noguchi T, et al. Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens. Nature 2014;515:577–81.
- [106] Carreno BM, Magrini V, Becker-Hapak M, Kaabinejadian S, Hundal J, Petti AA, et al. A dendritic cell vaccine increases the breadth and diversity ofmelanoma neoantigen-specific T cells. Science (80-) 2015;348:803–8.
- [107] Ott PA, Hu Z, Keskin DB, Shukla SA, Sun J, Bozym DJ, et al. An immunogenic personal neoantigen vaccine for patients with melanoma. Nature 2017;547:217–21.
- [108] Sahin U, Derhovanessian E, Miller M, Kloke B-P, Simon P, Löwer M, et al. Personalized RNA mutanome vaccines mobilize poly-specific therapeutic immunity against cancer. Nature 2017;547:222–6.
- [109] Keskin DB, Anandappa AJ, Sun J, Tirosh I, Mathewson ND, Li S, et al. Neoantigen vaccine generates intratumoral T cell responses in phase Ib glioblastoma trial. Nature 2019;565:234–9.
- [110] Hilf N, Kuttruff-Coqui S, Frenzel K, Bukur V, Stevanović S, Gouttefangeas C, et al. Actively personalized vaccination trial for newly diagnosed glioblastoma. Nature 2019;565:240–5.
- [111] Veatch JR, Jesernig BL, Kargl J, Fitzgibbon M, Lee SM, Baik C, et al. Endogenous CD4+ T Cells Recognize Neoantigens in Lung Cancer Patients, Including Recurrent Oncogenic KRAS and ERBB2 ( Her2) Driver Mutations. Cancer Immunol Res 2019;7:910–22. https://doi.org/10.1158/2326-6066.CIR-18-0402.
- [112] Tran E, Ahmadzadeh M, Lu Y, Gros A, Turcotte S, Robbins PF, et al. Immunogenicity of somatic mutations in human gastrointestinal cancers. Science (80-) 2015;350:1387–91.

- [113] Ren L, Leisegang M, Deng B, Matsuda T, Kiyotani K, Kato T, et al. Identification of neoantigenspecific T cells and their targets : implications for immunotherapy of head and neck squamous cell carcinoma. Oncoimmunology 2019;8:1–10. https://doi.org/10.1080/2162402X.2019.1568813.
- [114] Zhang X, Kim S, Hundal J, Herndon JM, Li S, Petti AA, et al. Breast Cancer Neoantigens Can Induce CD8 b T-Cell Responses and Antitumor Immunity. Cancer Immunol Res 2017;5:516–24. https://doi.org/10.1158/2326-6066.CIR-16-0264.
- [115] Nelde A, Walz JS, Kowalewski DJ, Wolz O, Peper JK, Gloria YC, et al. HLA class I-restricted MYD88 L265P-derived peptides as specific targets for lymphoma immunotherapy. Oncoimmunology 2017;6:1–11.

https://doi.org/10.1080/2162402X.2016.1219825.

- [116] Lo W, Parkhurst M, Robbins PF, Tran E, Lu Y-C, Jia L, et al. Immunologic Recognition of a Shared p53 Mutated Neoantigen in a Patient with Metastatic Colorectal Cancer. Cancer Immunol Res 2019;7:534– 43.
- [117] Yang W, Lee K-W, Srivastava RM, Kuo F, Krishna C, Chowell D, et al. Immunogenic neoantigens derived from gene fusions stimulate T cell responses. Nat Med 2019;25:767–75.
- [118] Cafri G, Yossef R, Pasetto A, Deniger DC, Lu Y, Parkhurst M, et al. Memory T cells targeting oncogenic mutations detected in peripheral blood of epithelial cancer patients. Nat Commun 2019;449:1–9. https://doi.org/10.1038/s41467-019-08304-z.
- [119] Malekzadeh P, Rosenberg SA, Drew C, Invest JC, Malekzadeh P, Pasetto A, et al. Neoantigen screening identifies broad TP53 mutant immunogenicity in patients with epithelial cancers. J Clin Invest 2019;129:1109–14.
- [120] Tran E, Turcotte S, Gros A, Robbins PF, Lu Y-C, Dudley ME, et al. Cancer Immunotherapy Based on Mutation-Specific CD4+ T Cells in a Patient with Epithelial Cancer. Science (80-) 2014;344:641–5.
- [121] Zacharakis N, Chinnasamy H, Black M, Xu H, Lu Y-C, Zheng Z, et al. Immune recognition of somatic mutations leading to complete durable regression in metastatic breast cancer. Nat Med 2018;24:724–30.
- [122] Chen F, Zou Z, Du J, Su S, Shao J, Meng F, et al. Neoantigen identification strategies enable personalized immunotherapy in refractory solid tumors. J Clin Invest 2019;129:2056–2070.
- [123] Sahin U, Türeci Ö. Personalized vaccines for cancer immunotherapy. Science (80-) 2018;359:1355–60.
- [124] Zitvogel L, Kepp O, Kroemer G. Essay Decoding Cell Death Signals in Inflammation and Immunity. Cell 2010;140:798–804. https://doi.org/10.1016/j.cell.2010.02.015.
- [125] Beatty GL, Gladney WL. Immune Escape Mechanisms as a Guide for Cancer Immunotherapy. Clin Cancer Res 2015;21:687–93. https://doi.org/10.1158/1078-0432.CCR-14-1860.
- [126] Zappasodi R, Merghoub T, Wolchok JD. Perspective Emerging Concepts for Immune Checkpoint Blockade-Based Combination Therapies.

### Biomedical Letters 2020; 6(1):48-59

Cancer Cell 2018;33:581–98. https://doi.org/10.1016/j.ccell.2018.03.005.

- [127] Patel SA, Minn AJ. Review Combination Cancer Therapy with Immune Checkpoint Blockade: Mechanisms and Strategies. Immunity 2018;48:417– 33. https://doi.org/10.1016/j.immuni.2018.03.007.
- [128] Galon J, Bruni D. Approaches to treat immune hot, altered and cold tumours with combination immunotherapies. Nat Rev Drug Discov 2019;18:197– 218.
- [129] Gotwals P, Cameron S, Cipolletta D, Cremasco V, Crystal A, Hewes B, et al. Prospects for combining targeted and conventional cancer therapy with immunotherapy. Nat Rev Cancer 2017;17:286–301.
- [130] Junjie W, J.Waxman D. Immunogenic chemotherapy: Dose and schedule dependence and combination with immunotherapy. Cancer Lett 2018;419:210–21.
- [131] Hu Z, Ott PA, Wu CJ. Towards personalized, tumour-specific, therapeutic vaccines for cancer. Nat Rev Immunol 2018;18:168–182.