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In silico elucidation of potent drug targets of the melanocyte-stimulating hormone receptor 1 protein

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Abstract

Most of the genes, including melanocortin 1 receptor (MC1R) involved in melanogenesis. The black to brown eumelanin and the yellow to red pheomelanin are the two kinds of pigment recognized in human skin. The primary cause of the dermal reaction to ultraviolet radiation (UVR) is cutaneous pigmentation, subsequently of the risk of developing skin cancer. Melanin pigmentation protects the skin against the harmful effects of ultraviolet radiation. The death rate of melanoma depends upon the type of skin cancer. Different computational approaches were utilized for 3D modeling and protein-protein docking analyses of MC1R leads to virtual screening. Comparative modeling and threading techniques were employed to predict the 3D structure of MC1R and 95.79% of quality factor was calculated. STRING database was utilized and 1Y7K was observed as interacting partner of MC1R. Protein-protein docking was performed, and potential interacting residues were observed. Virtual screening was performed against FDA library from ZINC database and molecular docking was performed by AutoDock Vina. The compound ZINC131 showed least binding energy of -10.3 kcal/mol. The suggested molecule may be used for further analyses in the drug discovery processes. In conclusion, the findings of current work may be helpful for novel therapeutic targets against MC1R.



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Introduction

The melanocortin receptor system has five receptors named as MC1R, MC2R, MC3R, MC4R, MC5R related to G-proteins and involved in the biological functions for multicellular animals and human beings [1]. Melanocyte-stimulating hormone (MSH) influenced through melanocyte receptors from melanoma. Such melanoma cells encode melanocyte-stimulating hormone receptor (MSHR), and coding DNA emulation also isolated from melanoma cells. The protein of 317 amino acids with transmembrane structural features of G-protein coupled receptors (GPCR) encoded from cloned coding DNA [2]. The melanin pigmentation can protect the skin from damaging possessions of ultraviolet radiation (UVR). In human skin, there are mainly two types of melanin, as red phaeomelanin and black eumelanin. Eumelanin has the potential to make free radicals against UVR. Hence, eumelanin is photo protective. The living beings with red hair having a prevalence of phaeomelanin in skin and hair and may abridged the aptitude to produce eumelanin. In mammals, MSH regulates a relative amount of phaeomelanin and eumelanin. MSH receptors increase the production of eumelanin [3, 4].

UVR may cause severe and prolonged effects on the skin of human beings. The result of ultraviolet (UV) radiation persuaded oxidative stress and defense through paracrine factors on the skin of living beings. The adaption in melanocytes due to heat stress influenced was reliant on melanin level and apoptosis reduction. The survival mechanism adopted by the role of δ -MSH in cell survival and melanocytes under ultraviolet oxidative stress [5]. About 65% of all melanomas are preventive measures from the protection of the sun. UVR is responsible for the appearance of all melanoma [6]. The primary gene present in humans is Melanocyte-Stimulating Hormone Receptor 1 (MC1R), where polymorphic coding regions show the foremost phenotypic variant factors in pigmentation. Though, distinct MC1R alleles influence the color of skin and hair [7]. MC1R member of GPCR may articulate in melanoma cells and MSH receptor. The extent and nature of major phenotypic variation factors may control by MSH receptor [8]. The phenotypic changes occur due to genes, mainly during clear Mendelian inheritance and the presence of no transitional phenotypes [9]. Some species including mouse, drosophila [10] and color carp [11] show pigmentation with the protagonist of mRNA or specific genes.

Furthermore, this gene plays an essential role in pigmentation leads to decisively phenotype and phenotype of beings affect due to gene interfaces [9], [10], [11], [12]. Due to pigmentation of MC1R, its variations may cause skin cancer [13]. MC1R has 80 variants. Out of these variants, some variants have partial loss of receptor signaling ability and enabled to produce cyclic adenosine monophosphate (cAMP) as the wild type receptor give active response to alpha-MSH stimulation. The red color phenotype belongs to the assessable change of melanin synthesis from eumelanin to phaeomelanin [14], [15]. The wide distribution of receptor is involved in several biological functions including pigmentation, antipyretic and anti-inflammatory actions [16].

The other variations in MC1R may increase the risk of evolving melanoma in the deficiency of UVR related to skin damage. Cancers are associated with mutations in the genes relate to melanoma risk including BRAF and CDKN2A. The activity of the receptor is mediated by GPCR which activate adenylate cyclase.

The primary purpose of this work was computational analyses, 3D structural prediction of MSHR, Protein-protein interaction studies and molecular docking analyses [17, 18]. For the completion of these objectives, sequence analysis, comparative modeling approach, comparative molecular docking approaches were utilized for analyses [19]. The result of trailed approaches confirmed that these strategies were accomplished of identifying the correspondent effective molecule.

Materials and Methods

The MC1R has the accession number Q01726 in Uniprot Knowledge Base. In present work, computational approaches including sequence analyses, 3D structure prediction, comparative protein-protein docking analyses and molecular docking analyses were performed.

ProtParam [20] and ProtScale [21] were used for the compositional properties analyses of the amino acid sequences.

The amino acid sequence of MC1R isoform was retrieved from Uniprot KB (<https://www.uniprot.org/uniprot/Q01726>) [22], and subjected to BLASTp against Protein Data Bank (PDB) [23] for the selection of suitable template identification. Homology modeling was done for 3D structure prediction of MC1R employed by using the automated protein modeling program MODELLER 9.20 [24]. Numerous other tools including SWISS MODEL [25], I-TASSER [26], PSIPred [27], rosetta

[28], Raptor-X [29], M4T [30], Phyre2 [31], SPARKS-X [32], IntFOLD [33], esyPred3D [34], HHpred [35], MOD-WEB [36], 3D-JigSaw [37] and CPHmodels [38] were utilized for 3D structure prediction. UCSF Chimera 1.13 [39] and PyMol [40] visualizing tools were used for the visualization of protein 3D structures. UCSF Chimera 1.13 was used for the minimization of predicted protein structure. For structure evaluation, MolProbity [41] online server was used. For the determination, the accuracy of protein sequence residues and protein structure quality, some other evaluation tools were used including Rampage [42], [43], Anolea [44], Errat [45], ProCheck [46] and Verify 3D [47]. Errat [48] evaluation tool was used for the structure prediction of the quality factor and Anolea was used to validate the Z-score value of the predicted structures.

STITCH [49] (Search Tool for InTeracting CHemical) [50] and STRING (Search Tool for the Retrieval of Interacting Genes/Protein) [51] database were used for the analyses of the functional interacting partner of MC1R [52].

Comparative molecular docking analyses were performed AutoDock Vina for library screening of FDA library of ZINC database for blind docking analyses. Through comparative molecular docking, the least energy of zinc compound was observed. The

molecular structure of the protein-ligand docking was visualized by UCSF Chimera 1.13.



Fig. 1: Location of MC1R on chromosome 16.

UCSC genome browser BLAT, Ensemble [58] and NCBI BLAST tools were used for the alignment of MC1R against the mouse and rat through Clustal Omega (**Fig. 2**).

COILS and ProtParam were employed to analyze the protein composition and ProtScale tools were used to calculate the composition of amino acids. It was observed that the molecular weight of the protein was 34705.51 based on the average isotope masses of amino acids [59] and the common isotope masses of amino acid and the common isotope mass of one water molecule. Theoretical pI which depends upon the side chain plays an important role in determining the pH of the protein. The half-life of the protein was observed 30 hours in vitro. The number of atoms having

Results and Discussion

Skin cancer is termed as Melanoma that reveals tumor heterogeneity features [53] and develops different research approaches as immunotherapy [53] [54] for the treatment and detection of skin cancer and melanoma risk [54]. The study of structural bioinformatics and oncology is the field of exploring information and efficiently providing research to understand better about melanoma. Research is providing better and possible resources for the treatment of skin cancer. Not only just reduction of melanoma risk or predictions of skin cancerous proteins are beneficial from the structural bioinformatics. The mechanism of MSHR protein role by using computational methods in neurological diseases and disorders [55] [56]. Vaccines also used for targeting melanoma tumor cells, as a form of active immunotherapy [54, 57].

Sequence Analyses

Many observations from the literature and examination of the biological database revealed information about the sequence MC1R. The UCSC genome browser was employed to examine the exact location of MC1R in human (**Fig. 1**).

negative and positive charges along with the total number of atoms and aliphatic index were also calculated (**Fig. 3**).

The neural network-based protein disorder was predicted in MC1R region and Pondr tool [60] was used to identify the natural disordered regions (**Fig. 4**).

Structure prediction

The 3D structure prediction of MC1R by using comparative modeling and threading approaches [61]. The NMR and X-ray crystallographic structure of MC1R was not reported. The sequence was submitted to BLASTp to retrieve the suitable

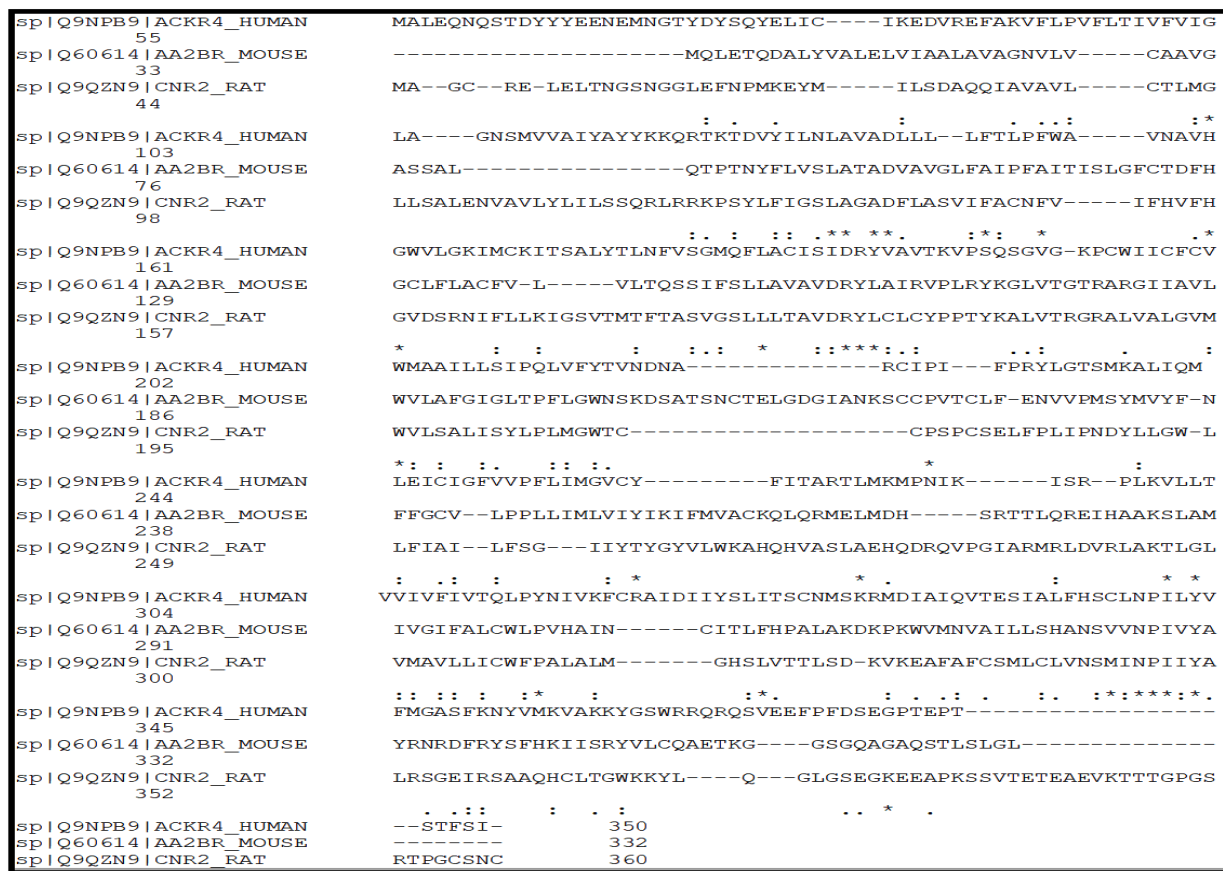


Fig. 2: Multiple sequence alignment through Clustal Omega against the sequences of mouse and rat showed conserved residues having *.

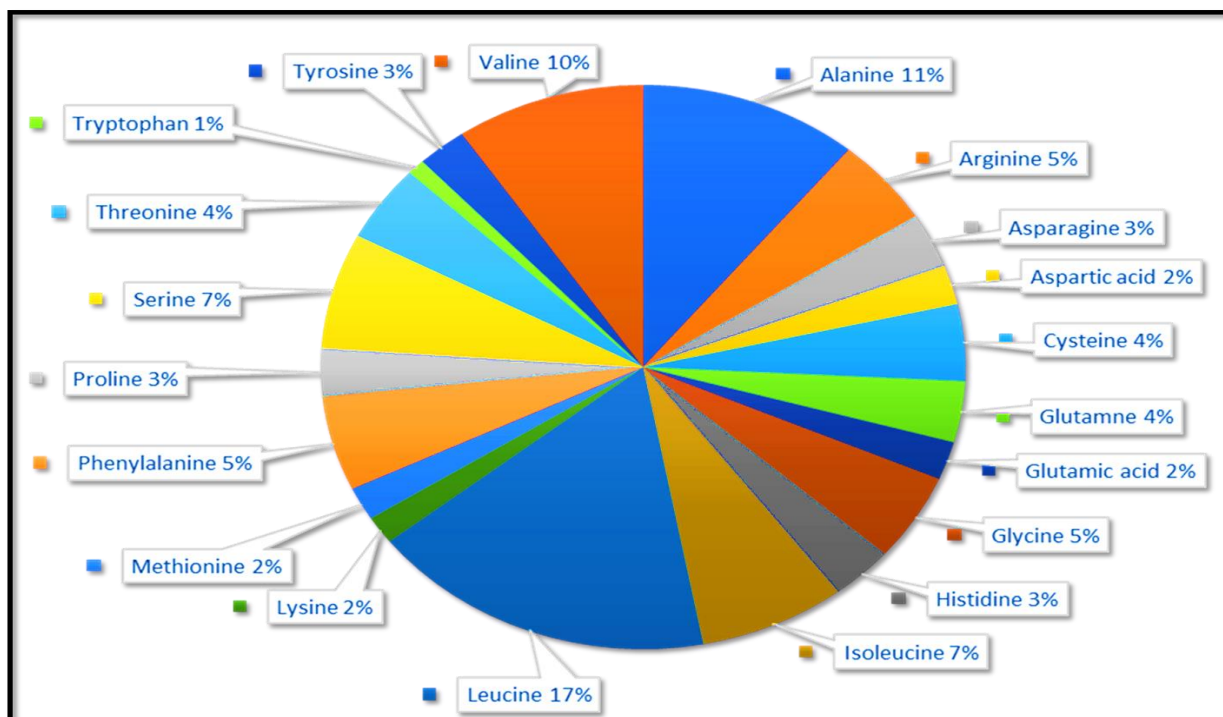


Fig. 3: Pie chart represents the composition of amino acid of MC1R and calculated percentage values.

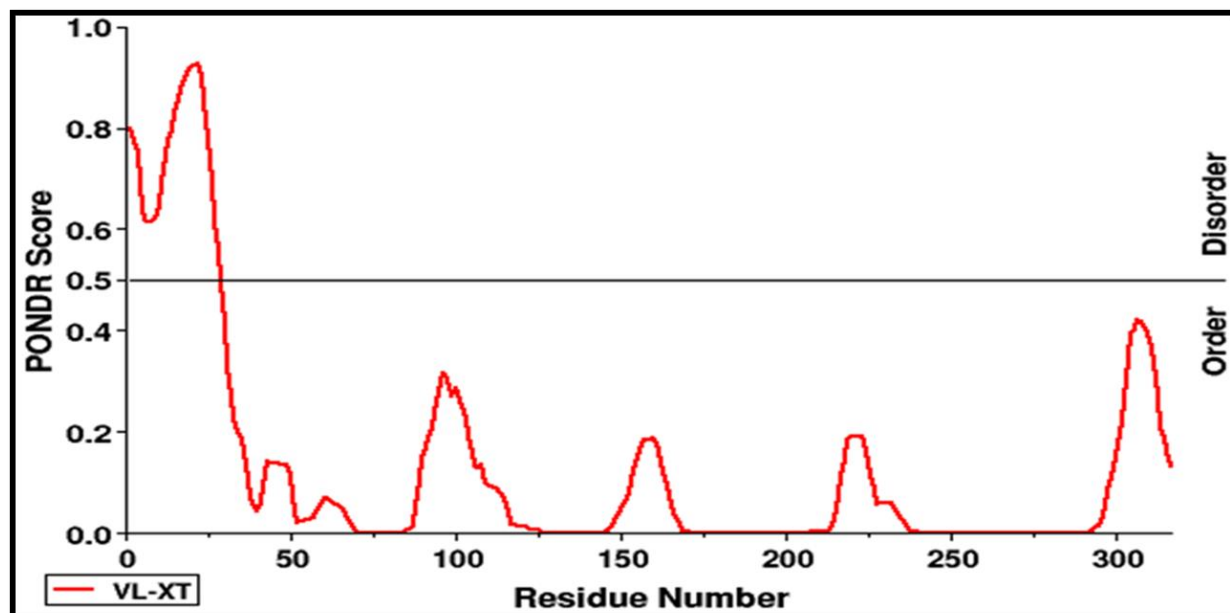


Fig. 4: Disorder Residue from PONDR tool of Protein.

templates against PDB. The top ranked five aligned suitable templates having maximum identity, query coverage, E values and total scores were selected for homology modeling. The selected templates were utilized to generate the 3D structure of MC1R. The overall query coverage and similarity for the used templates and MC1R protein showed 70% from end to end and was considered reliable for the homology modeling approach.

Several 3D models were predicted for MC1R by utilizing various tools through in silico approaches (threading and comparative modeling) to satisfy the sequence.

All the generated models were evaluated based on the quality factor, favored region, allowed region, and outliers. By using homology modeling and threading approaches, the graph was generated comparatively for all the predicted models and the reliable structure was retrieved from the generated graph (Fig. 5) for further analyses.

ERRAT evaluation tool was utilized and the observed overall quality factor of the selected structure of MC1R was 95.79%. For the evaluation of predicted models, the Ramachandran plot was utilized which reveals the ϕ and ψ distributed and 99.3% residues were observed in favored region and allowed region and only 1 residue was observed in outlier region. For the improvement of stereochemistry, the structural minimization on the predicted structure was applied and considered the model for the most optimal purpose. By using UCSF Chimera 1.13, the most optimal structure was

minimized on 1000 steepest and 1000 conjugates gradients run after the critical examination at evaluation parameters (Fig. 6).

Protein-Protein Interactions

The protein-protein docking analyses were performed and determined by using GramX online server [62]. The interacting residues of the complex were visualized through UCSF Chimera 1.13 (Fig. 7) (Table 1).

Comparative Molecular Docking

Different binding energies and complexes were revealed by comparative molecular docking analyses. The best complex was determined by analyzing the least binding energy and selected for the interactional studies. The compound ZINC00131 showed least binding energy of -10.4 kcal/mol (Table 2). The 3D structure analyses of comparative molecular docking was visualized by UCSF Chimera 1.13 (Fig. 8) [66]. The 2D structure for the compound was drawn by ChemDrawUltra 8.0 [65] (Fig. 9).

Proteins are involved in cellular processes and are the molecule of life. The functionality of proteins depends on its structure. The field of bioinformatics covered various disciplines including structural bioinformatics, computing, mathematics, artificial intelligence, computational chemistry and biostatistics approaches to facilitate the discovery of

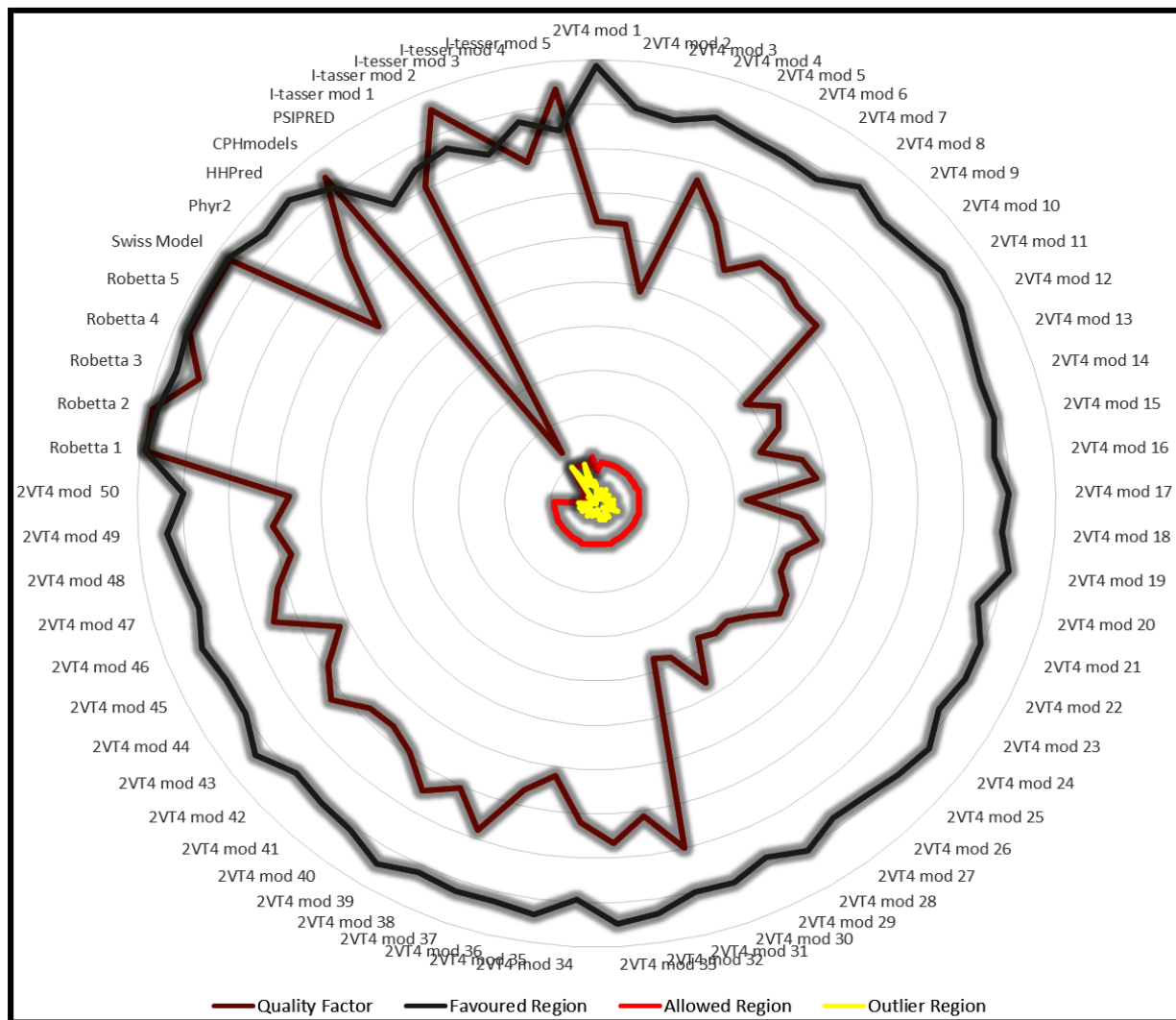


Fig. 5: Graph of quality factor, favored region, allowed region and outliers.

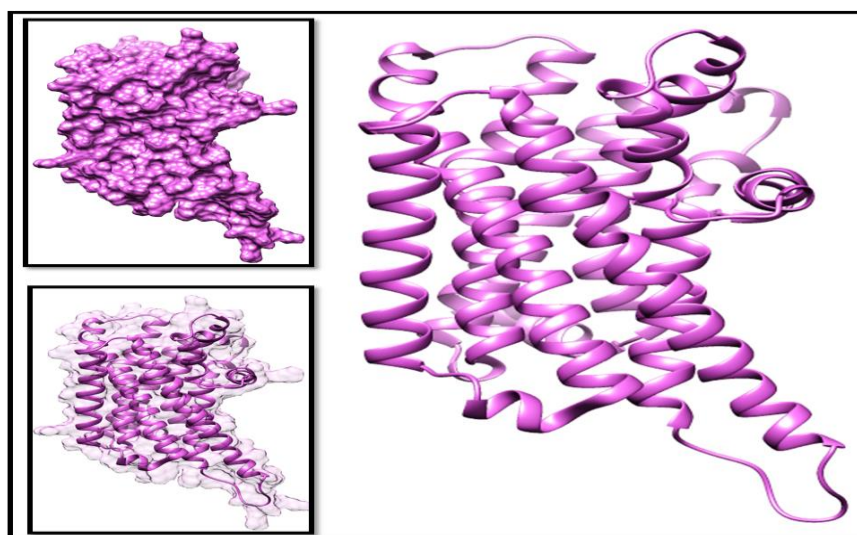


Fig. 6: 3D predicted structure of MC1R.

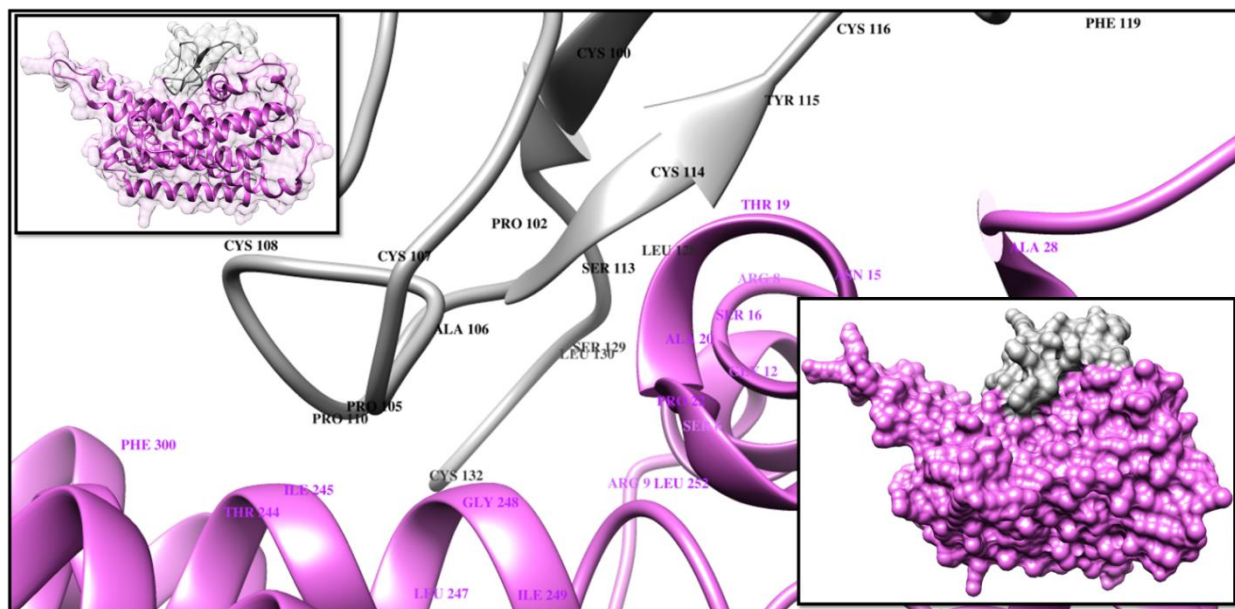


Fig. 7: Protein-protein docking analyses.

Table 1: Residues of protein-protein docking as receptor protein and ligand protein.

Targeting Protein Name	Targeting Protein residues	Interacting Protein Name	Interacting Protein residues
Melanocyte-Stimulating Hormone Receptor 1	PHE 300, THR 244, ILE 245, GLY 248, LEU 247, ILE 249, ARG 9, LEU 252, SER 6, PRO 22, GLY 12, ALA 20, SER 16, ARG 8, ASN 15, ALA 28,	1Y7K	CYS 108, PRO 110, PRO 105, CYS 107, ALA 106, CYS 132, PRO 102, LEU 130, SER 129, SER 113, LEU 128, CYS 114, CYS 100, TYR 115, CY

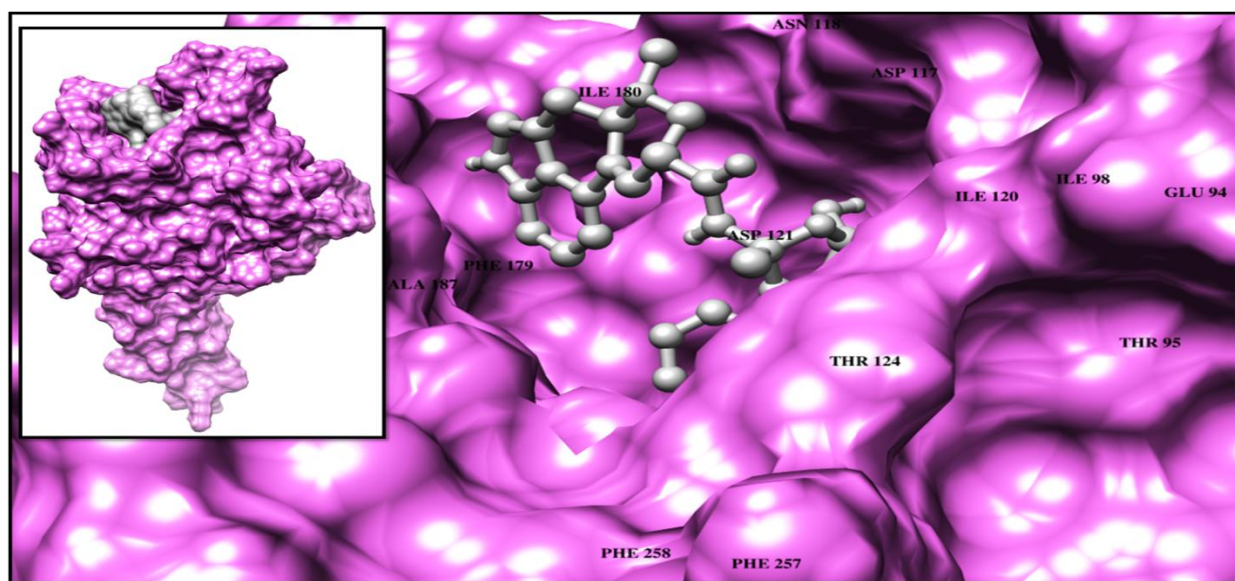


Fig. 8: Molecular docking analyses of ZINC131 compound having least binding energy.

Table 2: Top four least binding energy compounds from molecular docking analyses.

Compound	Binding Affinity kcal/mol	RMSD/ upper binding	RMSD/ Lower binding
ZINC00131	-10.0	0	0
ZINC00087	-9.7	0	0
ZINC00059	-9.5	0	0
ZINC00092	-9.4	0	0

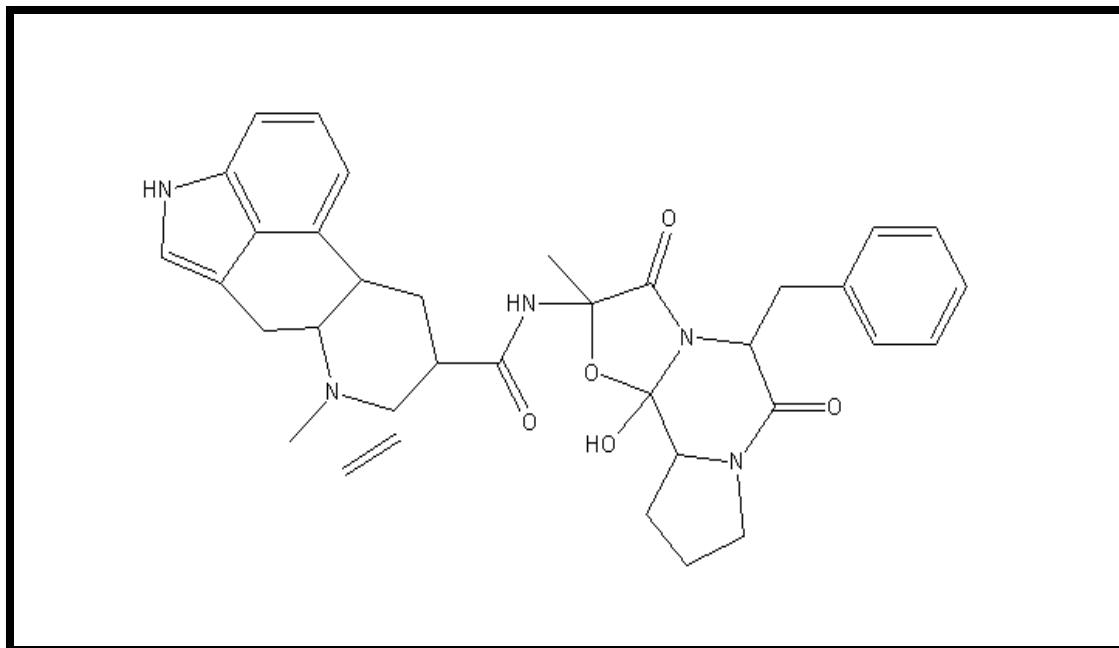


Fig. 9: 2D structure of ZINC131 compound.

new biological ideas [67]. Over the last ten years, the structural bioinformatics has undergone many improvements. Biological data increases with the help of computational resources and methodology to develop the size and resolution of study as well as created complex questions to research [68, 69]. The approaches of computational analyses lower the time phases and very useful to the researcher in the field of research [70]. By using *in silico* methods and computational approaches predict the protein structure of MC1R.

Conclusion

Various computational approaches were applied for the structure prediction of MC1R helped to develop novel drug targets. Protein-protein docking analyses were performed and interactional residues were reported. Comparative molecular docking was utilized and ZINC131 compounds showed least binding energy.

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Conflicts of Interest

The authors declare no conflict of Interest.

References

- [1] Cai M, Hruby VJ. The Melanocortin Receptor System: A Target for Multiple Degenerative Diseases. *Curr Protein Pept Sci* 2016;17(5):488-96.
- [2] Chhajlani V, Wikberg JE. Molecular cloning and expression of the human melanocyte stimulating hormone receptor cDNA. *FEBS Lett* 1992;309(3):417-20.
- [3] Valverde P, Healy E, Jackson I, Rees JL, Thody AJ. Variants of the melanocyte-stimulating hormone receptor gene are associated with red hair and fair skin in humans. *Nature Genetics* 1995;11(3):328-330.
- [4] Natarajan VT, Ganju P, Ramkumar A, Grover R, Gokhale RS. Multifaceted pathways protect human skin from UV radiation. *Nat Chem Biol* 2014;10(7):542-51.
- [5] Del Bino S, Duval C, Bernerd F. Clinical and Biological Characterization of Skin Pigmentation Diversity and Its Consequences on UV Impact. *Int J Mol Sci* 2018;19(9).
- [6] Emri G, Paragh G, Tosaki A, Janka E, Kollar S, Hegedus C, Gellen E, Horkay I, Koncz G, Remenyik E. Ultraviolet radiation-mediated development of cutaneous melanoma: An update. *J Photochem Photobiol B* 2018;185:169-175.
- [7] Duffy DL, Box NF, Chen W, Palmer JS, Montgomery GW, James MR, Hayward NK, Martin NG, Sturm RA. Interactive effects of MC1R and OCA2 on melanoma risk phenotypes. *Hum Mol Genet* 2004;13(4):447-61.

- [8] Garcia-Borron JC, Sanchez-Laorden BL, Jimenez-Cervantes C. Melanocortin-1 receptor structure and functional regulation. *Pigment Cell Res* 2005;18(6):393-410.
- [9] Dong C, Wang H, Xue L, Dong Y, Yang L, Fan R, Yu X, Tian X, Ma S, Smith GW. Coat color determination by miR-137 mediated down-regulation of microphthalmia-associated transcription factor in a mouse model. *RNA* 2012;18(9):1679-86.
- [10] Kennell JA, Cadigan KM, Shakhmantsir I, Waldron EJ. The MicroRNA miR-8 is a positive regulator of pigmentation and eclosion in *Drosophila*. *Developmental Dynamics* 2012;241(1):161-168.
- [11] Chen H, Wang J, Du J, Si Z, Yang H, Xu X, Wang C. ASIP disruption via CRISPR/Cas9 system induces black patches dispersion in Oujiang color common carp. *Aquaculture* 2019;498:230-235.
- [12] Mandal BK, Chen H, Si Z, Hou X, Yang H, Xu X, Wang J, Wang C. Shrunk and scattered black spots turn out due to MC1R knockout in a white-black Oujiang color common carp (*Cyprinus carpio* var. color). *Aquaculture* 2020;518:734822.
- [13] Gerstenblith MR, Goldstein AM, Fagnoli MC, Peris K, Landi MT. Comprehensive evaluation of allele frequency differences of MC1R variants across populations. *Human mutation* 2007;28(5):495-505.
- [14] Raimondi S, Sera F, Gandini S, Iodice S, Caini S, Maisonneuve P, Fagnoli MC. MC1R variants, melanoma and red hair color phenotype: a meta-analysis. *Int J Cancer* 2008;122(12):2753-60.
- [15] Schiöth HB, Phillips SR, Rudzish R, Birch-Machin MA, Wikberg JE, Rees JL. Loss of function mutations of the human melanocortin 1 receptor are common and are associated with red hair. *Biochemical and biophysical research communications* 1999;260(2):488-491.
- [16] Cooray SN, Clark AJ. Melanocortin receptors and their accessory proteins. *Molecular and cellular endocrinology* 2011;331(2):215-221.
- [17] By Sheikh Arslan Sehgal AHM, Rana Adnan Tahir, Asif Mir. *Quick Guideline for Computational Drug Design*. Bentham Science Publisher 2018.
- [18] Sehgal SA. *Pharmacoinformatics, Adaptive Evolution, and Elucidation of Six Novel Compounds for Schizophrenia Treatment by Targeting DAOA (G72) Isoforms*. *Biomed Res Int* 2017;2017:5925714.
- [19] Tahir RA, Sehgal SA. *Pharmacoinformatics and Molecular Docking Studies Reveal Potential Novel Compounds Against Schizophrenia by Target SYN II*. *Comb Chem High Throughput Screen* 2018;21(3):175-181.
- [20] Goud TS, Upadhyay RC, Onteru SK, Pichili VBR, Chadipiralla K. Identification and sequence characterization of melanocortin 1 receptor gene (MC1R) in *Bos indicus* versus (*Bos taurus* X *Bos indicus*). *Animal biotechnology* 2019:1-12.
- [21] Gasteiger E, Hoogland C, Gattiker A, Wilkins MR, Appel RD, Bairoch A. Protein identification and analysis tools on the ExPASy server. *The proteomics protocols handbook*: Springer; 2005. p. 571-607.
- [22] Boutet E, Lieberherr D, Tognolli M, Schneider M, Bairoch A. Uniprotkb/swiss-prot. *Plant bioinformatics*: Springer; 2007. p. 89-112.
- [23] Rose PW, Prlić A, Altunkaya A, Bi C, Bradley AR, Christie CH, Costanzo LD, Duarte JM, Dutta S, Feng Z. The RCSB protein data bank: integrative view of protein, gene and 3D structural information. *Nucleic acids research* 2016:gkw1000.
- [24] Webb B, Sali A. *Protein structure modeling with MODELLER*. *Protein structure prediction*: Springer; 2014. p. 1-15.
- [25] Arnold K, Bordoli L, Kopp J, Schwede T. The SWISS-MODEL workspace: a web-based environment for protein structure homology modelling. *Bioinformatics* 2006;22(2):195-201.
- [26] Zhang Y. I-TASSER server for protein 3D structure prediction. *BMC bioinformatics* 2008;9(1):40.
- [27] Zhang H, Tang H, Zhang L, Jin L, Tang Y. Evaluation on prediction methods of protein secondary structure. *Computers and Applied Chemistry* 2003;20(6):735-740.
- [28] Kim DE, Chivian D, Baker D. Protein structure prediction and analysis using the Robetta server. *Nucleic acids research* 2004;32(suppl_2):W526-W531.
- [29] Källberg M, Wang H, Wang S, Peng J, Wang Z, Lu H, Xu J. Template-based protein structure modeling using the RaptorX web server. *Nature protocols* 2012;7(8):1511.
- [30] Fernandez-Fuentes N, Madrid-Aliste CJ, Rai BK, Fajardo JE, Fiser A. M4T: a comparative protein structure modeling server. *Nucleic acids research* 2007;35(suppl_2):W363-W368.
- [31] Kelley LA, Mezulis S, Yates CM, Wass MN, Sternberg MJ. The Phyre2 web portal for protein modeling, prediction and analysis. *Nature protocols* 2015;10(6):845.
- [32] Yang Y, Faraggi E, Zhao H, Zhou Y. Improving protein fold recognition and template-based modeling by employing probabilistic-based matching between predicted one-dimensional structural properties of query and corresponding native properties of templates. *Bioinformatics* 2011;27(15):2076-2082.
- [33] McGuffin LJ, Adiyaman R, Maghrabi AH, Shuid AN, Brackenridge DA, Nealon JO, Philomina LS. IntFOLD: an integrated web resource for high performance protein structure and function prediction. *Nucleic acids research* 2019;47(W1):W408-W413.
- [34] Lambert C, Leonard N, De Bolle X, Depiereux E. ESyPred3D: Prediction of proteins 3D structures. *Bioinformatics* 2002;18(9):1250-1256.
- [35] Zimmermann L, Stephens A, Nam S-Z, Rau D, Kübler J, Lozajic M, Gabler F, Söding J, Lupas AN, Alva V. A completely reimplemented MPI bioinformatics toolkit with a new HHpred server at its core. *Journal of molecular biology* 2018;430(15):2237-2243.
- [36] Pieper U, Webb BM, Dong GQ, Schneidman-Duhovny D, Fan H, Kim SJ, Khuri N, Spill YG, Weinkam P, Hammel M. ModBase, a database of annotated comparative protein structure models and associated resources. *Nucleic acids research* 2014;42(D1):D336-D346.

- [37] Bates PA, Kelley LA, MacCallum RM, Sternberg MJ. Enhancement of protein modeling by human intervention in applying the automatic programs 3D-JIGSAW and 3D-PSSM. *Proteins: Structure, Function, and Bioinformatics* 2001;45(S5):39-46.
- [38] Nielsen M, Lundegaard C, Lund O, Petersen TN. CPHmodels-3.0—remote homology modeling using structure-guided sequence profiles. *Nucleic acids research* 2010;38(suppl_2):W576-W581.
- [39] Pettersen EF, Goddard TD, Huang CC, Couch GS, Greenblatt DM, Meng EC, Ferrin TE. UCSF Chimera—a visualization system for exploratory research and analysis. *Journal of computational chemistry* 2004;25(13):1605-1612.
- [40] Bramucci E, Paiardini A, Bossa F, Pascarella S. PyMod: sequence similarity searches, multiple sequence-structure alignments, and homology modeling within PyMOL. *BMC bioinformatics* 2012;13(S4):S2.
- [41] Chen VB, Arendall WB, Headd JJ, Keedy DA, Immormino RM, Kapral GJ, Murray LW, Richardson JS, Richardson DC. MolProbity: all-atom structure validation for macromolecular crystallography. *Acta Crystallographica Section D: Biological Crystallography* 2010;66(1):12-21.
- [42] Lovell SC, Davis IW, Arendall III WB, De Bakker PI, Word JM, Prisant MG, Richardson JS, Richardson DC. Structure validation by α geometry: ϕ , ψ and $\text{C}\beta$ deviation. *Proteins: Structure, Function, and Bioinformatics* 2003;50(3):437-450.
- [43] Wang W, Xia M, Chen J, Deng F, Yuan R, Zhang X, Shen F. Data set for phylogenetic tree and RAMPAGE Ramachandran plot analysis of SODs in *Gossypium raimondii* and *G. arboreum*. *Data in brief* 2016;9:345-348.
- [44] Melo F, Devos D, Depiereux E, Feytmans E, editors. ANOLEA: a www server to assess protein structures. *Ismb*; 1997.
- [45] Dym O, Eisenberg D, Yeates T. ERRAT. 2012.
- [46] Laskowski RA, MacArthur MW, Moss DS, Thornton JM. PROCHECK: a program to check the stereochemical quality of protein structures. *Journal of applied crystallography* 1993;26(2):283-291.
- [47] Grotthuss Mv, Pas J, Wyrwicz L, Ginalski K, Rychlewski L. Application of 3D-Jury, GRDB, and Verify3D in fold recognition. *Proteins: Structure, Function, and Bioinformatics* 2003;53(S6):418-423.
- [48] Colovos C, Yeates TO. Verification of protein structures: patterns of nonbonded atomic interactions. *Protein science* 1993;2(9):1511-1519.
- [49] Kuhn M, von Mering C, Campillos M, Jensen LJ, Bork P. STITCH: interaction networks of chemicals and proteins. *Nucleic acids research* 2007;36(suppl_1):D684-D688.
- [50] Kuhn M, Szklarczyk D, Pletscher-Frankild S, Blicher TH, Von Mering C, Jensen LJ, Bork P. STITCH 4: integration of protein–chemical interactions with user data. *Nucleic acids research* 2014;42(D1):D401-D407.
- [51] Franceschini A, Szklarczyk D, Frankild S, Kuhn M, Simonovic M, Roth A, Lin J, Minguez P, Bork P, Von Mering C. STRING v9. 1: protein-protein interaction networks, with increased coverage and integration. *Nucleic acids research* 2012;41(D1):D808-D815.
- [52] A. Sehgal SAFK, Seemab. In Silico Analyses, Bioequivalence and Disposition Kinetics of Allopurinol in Healthy Male Subjects. *Drug Delivery Letters* 2016;6:113-121.
- [53] Grzywa TM, Paskal W, Włodarski PK. Intratumor and intertumor heterogeneity in melanoma. *Translational oncology* 2017;10(6):956-975.
- [54] Rodríguez-Cerdeira C, Carnero Gregorio M, López-Barcenas A, Sánchez-Blanco E, Sánchez-Blanco B, Fabbrocini G, Bardhi B, Sinani A, Guzman RA. Advances in immunotherapy for melanoma: a comprehensive review. *Mediators of inflammation* 2017;2017.
- [55] Ding Z, Kihara D. Computational Methods for Predicting Protein-Protein Interactions Using Various Protein Features. *Current protocols in protein science* 2018;93(1):e62.
- [56] Leuschner C, Alila H. Methods for diagnosis of and predicting treatment efficacy of hormone receptor expressing tumors, cancers and neoplasias. *Google Patents*; 2019.
- [57] Engelstein R, Merims S, Eisenberg G, Cohen J, Frank S, Hamburger T, Frankenburg S, Ron I, Isacson R, Grenader T. Immune monitoring of patients treated with a whole-cell melanoma vaccine engineered to express 4-1BBL. *Journal of Immunotherapy* 2016;39(8):321-328.
- [58] Hubbard T, Barker D, Birney E, Cameron G, Chen Y, Clark L, Cox T, Cuff J, Curwen V, Down T. The Ensembl genome database project. *Nucleic acids research* 2002;30(1):38-41.
- [59] Dietzen DJ. Amino acids, peptides, and proteins. *Principles and Applications of Molecular Diagnostics*: Elsevier; 2018. p. 345-380.
- [60] Djulbegovic MB, Uversky VN. Expanding the understanding of the heterogeneous nature of melanoma with bioinformatics and disorder-based proteomics. *International journal of biological macromolecules* 2020;150:1281-1293.
- [61] Emamjomeh A, Choobineh D, Hajieghrari B, MahdiNezhad N, Khodavirdipour A. DNA–protein interaction: identification, prediction and data analysis. *Molecular biology reports* 2019:1-26.
- [62] Tovchigrechko A, Vakser IA. GRAMM-X public web server for protein–protein docking. *Nucleic acids research* 2006;34(suppl_2):W310-W314.
- [63] Tahir RA, Hassan F, Kareem A, Iftikhar U, Sehgal SA. Ligand-Based Pharmacophore Modeling and Virtual Screening to Discover Novel CYP1A1 Inhibitors. *Curr Top Med Chem* 2019;19(30):2782-2794.
- [64] Tahir RA, Wu H, Rizwan MA, Jafar TH, Saleem S, Sehgal SA. Immunoinformatics and molecular docking studies reveal potential epitope-based peptide vaccine against DENV-NS3 protein. *J Theor Biol* 2018;459:162-170.
- [65] Arshad M. Heterocyclic compounds bearing pyrimidine, oxazole and pyrazole moieties: design, computational, synthesis, characterization, antibacterial and molecular docking screening. *SN Applied Sciences* 2020;2(3):1-8.

- [66] Sehgal SA. Pharmacoinformatics and molecular docking studies reveal potential novel Proline Dehydrogenase (PRODH) compounds for Schizophrenia inhibition. *Medicinal Chemistry Research* 2017;26:314-326.
- [67] Yang X, Wang Y, Byrne R, Schneider G, Yang S. Concepts of artificial intelligence for computer-assisted drug discovery. *Chemical reviews* 2019;119(18):10520-10594.
- [68] McQuin C, Goodman A, Chernyshev V, Kamensky L, Cimini BA, Karhohs KW, Doan M, Ding L, Rafelski SM, Thirstrup D. CellProfiler 3.0: Next-generation image processing for biology. *PLoS biology* 2018;16(7).
- [69] Tahir RA, Bashir A, Yousaf MN, Ahmed A, Dali Y, Khan S, Sehgal SA. In Silico identification of angiotensin-converting enzyme inhibitory peptides from MRJPI. *PLoS One* 2020;15(2):e0228265.
- [70] Cristino TM, Neto AF, Costa AFB. Energy efficiency in buildings: analysis of scientific literature and identification of data analysis techniques from a bibliometric study. *Scientometrics* 2018;114(3):1275-1326.